



INTERNATIONAL JOURNAL
OF
PHYTOPHARMACY RESEARCH
www.phytopharmacyresearch.com

PHYTOCHEMICAL SCREENING AND ANTIOXIDANT ACTIVITY OF *LEPIDIUM SATIVUM* LINN. SEEDS

Anu S, Deepashree GH Math, Shruthi S, Vandana Singh, Shivaji Bole*, Sam Balu,
Vedamurthy AB

Biotechnology Finishing School, Dept. of Biotechnology, The Oxford College of Science,
19thMain, 17th B Cross, HSR layout, Bangalore.

ABSTRACT

The objective of this study is to investigate the phytochemical constituents. Phytochemical screening of seeds of *Lepidium sativum linn* (family-Brassicaceae) in different organic solvent revealed the presence of alkaloids, tannins, flavonoids. Antioxidant activities were evaluated by using *in-vitro* antioxidant assay models like phospho-molybdenum and reducing power assay. And hydrogen peroxide assay. The percentage of antioxidant activity by phospho-molybdenum assay was in the order methanol>ethanol>acetone. The reducing power assay was maximum shown in order acetone>methanol>ethanol and hydrogen peroxide assay was found in order acetone>ethanol>methanol. The results obtained in this study showed that the seeds of *lepidium sativum linn* have antioxidative properties which provide a basis for the traditional use of the plant.

Keywords: *Lepidium sativum linn*, Reducing power assay, H₂O₂ scavenging.

INTRODUCTION

Lepidium sativum linn (Garden cress) is an annual herb, belonging to Brassicaceae family. It is a fast-growing, edible plant botanically related to watercress and mustard and sharing their peppery, tangy flavor and aroma. Seeds, leaves and roots are economically important, however, the crop is mainly cultivated for seeds. *Lepidium sativum linn* in India and it is an important medicinal crop in India [1-3].



The seeds are useful both, internally as well as externally. The paste of seeds is applied in rheumatic joints to relieve the pain and swelling. The pulp of the seeds applied on the painful joints and on haematoma, effectively reduces the pain and swelling. The paste is salutary in the treatment of various skin diseases also.

Reactive oxygen species (ROS) are a class of highly reactive molecules derived from the metabolism of oxygen. ROS, including superoxide radicals, hydroxyl

radical and hydrogen peroxide molecules are often generated as by-products of biological reactions or from exogenous factors. There is extensive evidence to involve ROS in the development of degenerative diseases. Evidence suggests that compounds especially from natural sources are capable of providing protection against free radicals. This has attracted a great deal of research interest in natural antioxidants. It is necessary to Screen out medicinal plants for their antioxidant potential [4-7].

MATERIALS AND METHODS

Plant Collection

Seeds of *L.sativum linn* were collected from Hubli, Karnataka and authenticated by Botanist, Bangalore University, Bangalore, Karnataka, India.

Chemicals and Reagents

All the chemicals and reagents used were of analytical grade and are purchased from Lancaster Research Lab, Chennai, India and Hi-media Lab, Mumbai, India.

EXTRACTION OF PLANT MATERIAL

Shade dried seed of *L.sativum linn* was powdered and about 15g of shade dried powder of seeds of *lepidium sativum linn* was successively extracted with

ethanol, acetone, and methanol. The extract obtained was filtered, concentrated by evaporating at 100°C in a water bath and dried [8-10].

PHYTOCHEMICAL ANALYSIS

The extracts were used for preliminary screening of phytochemicals such as alkaloids (Wagner's and Meyer's tests), saponins (foam and froth tests), (Acetone-water test), FeCl₃ test, tannins (gelatin test), and flavonoids (Alkaline reagent and Lead acetate tests), the screening was done as per the standard method.

Test for alkaloids

- Dragendorff's test: 2mg of the test extract and 5ml of distilled water was added, 2M hydrochloric acid was added until an acid reaction occurs. To this 1mL of Dragendorff's reagent was added. Formation of orange red precipitate indicated the presence of alkaloids.
- Hager's test: 2mg of the test extract was taken in a test-tube, a few drops of Hager's reagent was added. Formation of yellow precipitate confirmed the presence of alkaloids.
- Wagner's test: 2mg of extract was acidified with 1.5% v/v of hydrochloric acid and a few drops of Wagner's reagent were added. A yellow or brown precipitate indicated the presence of alkaloids.
- Mayer's test: few drops of Mayer's reagent, 2mg of extract was added formation of white or pale yellow precipitation indicated the presence of alkaloids [11].

Test for flavonoids

- Ferric chloride test: test solution with few drops of ferric chloride solution shows intense green color
- Zinc hydrochloride acid reduction test: test solution with zinc dust and few drops of hydrochloric acid shows magenta red color
- Lead acetate solution test: test solution with few drops of lead acetate (10%) solution gives yellow precipitate [12].

Tests for saponins

- Foam test: to the extract solution, a drop of sodium bicarbonate solution was added. The test tube was shaken vigorously and left for 3 minutes. The formation of honeycomb like froth indicates the presence of saponins.

Test for steroids

- Salkowski reaction: 2mg of dry test extract was shaken with chloroform, to the chloroform layer sulphuric acid was added slowly by the sides of the test tube. Formation of red colour indicated the presence of steroids

Test for tannins

- Ferric chloride test: 1-2ml of the extract, few drops of 5% w/v FeCl₃ solution was added. A green colour indicates the presence of gallotannins; while brown colour indicates the presence of pseudotannins.
- Gelatine test: test solution when treated with gelatine solution gives white precipitate

Test for glycosides

- Keller-killiani test: the test solution was treated with few drops of ferric chloride solution and mixed. When conc. Sulphuric acid containing ferric chloride solution was added, it forms two layers, lower layer reddish brown and the upper layer acetic acid layer turns bluish green
- Bromine water test: test solution when dissolved in bromine water gives yellow precipitate

ANTIOXIDANT ACTIVITY

Determination of phospho-molybdenum assay

The antioxidant activity of all the extract was determined by the phospho-molybdenum Method as described by Prieto *et al* [13]. 0.3 ml of extract of different concentrations (100 to 500 µg/ml) was mixed with 3 ml of reagent solution (0.6 M sulfuric acid, 28 mM sodium phosphate and 4 mM ammonium molybdate). The reaction mixture was incubated at 95°C for 90 min and cooled to room temperature. Finally, absorbance was measured at 695 nm using a spectrophotometer against blank. Distilled water (0.3 ml) in place of extract was used as the blank. The total antioxidant capacity was expressed as the number of equivalents of Ascorbic acid.

REDUCING POWER ASSAY (IRON (III) TO IRON (II) REDUCTION)

Reducing power is associated with antioxidant activity and may serve as a significant reflection of the antioxidant activity. Compounds with reducing power indicate that they are electron donors and can reduce the oxidized intermediates of lipid peroxidation processes, so that they can act as primary and secondary antioxidants

The Ferric reducing power method was applied with slight modifications of the method in which, 2.5 mL of extract solution of different concentrations (100 to 500 µg/mL) was mixed with phosphate buffer (2.5 mL, 0.2 M, pH 6.6) and potassium ferricyanide (2.5 mL, 0.1%). This was incubated at 50°C for 20 min. After the incubation, 2.5 mL of 10% trichloroacetic acid was added. 2.5 mL of the reaction mixture was mixed with distilled water (2.5 mL) and ferric chloride (0.5 mL, 0.1%). The absorbance was measured at 700 nm. Increasing absorbance of the reaction mixture indicates increasing reducing power. The same procedure was applied for ascorbic acid which acts as the standard. Increase in the absorbance indicates increase in reducing power.

H₂O₂ SCAVENGING ASSAY

Hydrogen peroxide occurs naturally at low concentration levels in the air, water, human body, plants, microorganisms, food and beverages. It is widely used as a bleaching agent in the textile, paper and pulp industries. Human beings exposed to H₂O₂ indirectly via the environment are estimated as 0.28 mg/kg/day with intake from leaf crops contributing most to this exposure. Hydrogen peroxide enters the human body through inhalation of vapour or mist and through eye or skin contact. In the body, H₂O₂ is rapidly decomposed into oxygen and water and this may produce hydroxyl radicals

(OH[·]) that can initiate lipid peroxidation and cause DNA damage [14-15].

A solution of hydrogen peroxide (40 mM) is prepared in phosphate buffer (50mM pH 7.4). The concentration of hydrogen peroxide is determined by absorption at 230 nm using a spectrophotometer. Extract (100-1000mg/ml) in distilled water is added to hydrogen peroxide and absorbance at 230 nM is determined after 10 min against a blank solution containing phosphate buffer without hydrogen peroxide. The percentage of hydrogen peroxide scavenging is calculated as follows:

$$\% \text{ scavenged (H}_2\text{O}_2) = (\text{Abs control} - \text{Abs standard} / \text{Abs control}) \times 100$$

RESULT AND DISCUSSION

In this study of *In Vitro* antioxidant activity, preliminary phytochemical screening of the different fractions of seed extracts revealed that the presence of

alkaloids, flavonoids, saponins, steroid, tannins and glycosides and the results are tabulated in Table 1. The results of the free radical scavenging potentials of different fractions tested by reducing power assay method Phospho-molybdenum assay and hydrogen peroxide assay are depicted in fig 1, 2 and 3.

The phospho-molybdate method is quantitative, since the total antioxidant capacity is expressed as ascorbic acid equivalents. The percentage of antioxidant activity was in the order methanol>ethanol>acetone.

In reducing power assay, acetone fraction seemed to have quite high reducing activity compared to methanol and ethanol. The reducing power assay of different extract is given in fig 2.

In hydrogen peroxide assay, acetone extract shows the highest scavenging activity followed by ethanol and methanol extract when compared to the standard ascorbic acid which is depicted in fig 3.

Table 1. Screening of Secondary metabolites

	Ethanol	Methanol	Acetone
Test for alkaloids			
Dragendorff's test	-	+	-
Hager's test	+	+	+
Mayer's test	+	+	+
Wagner's test	+	+	-
Test for flavonoids			
FeCl ₃ test	-	+	+
Zinc test	-	-	-
Alkaline reagent test	+	+	+
Test for saponins			
Foam test	-	+	-
Test for tannins			
Gelatine test	-	+	-
Ferric chloride test	-	+	+
Test for steroids			
Salkowaski reaction	-	+	+
Test for glycosides			
Keller killiani test	-	+	+
Bromine water test	-	+	-

Fig 1. Different fractions tested by reducing power assay method

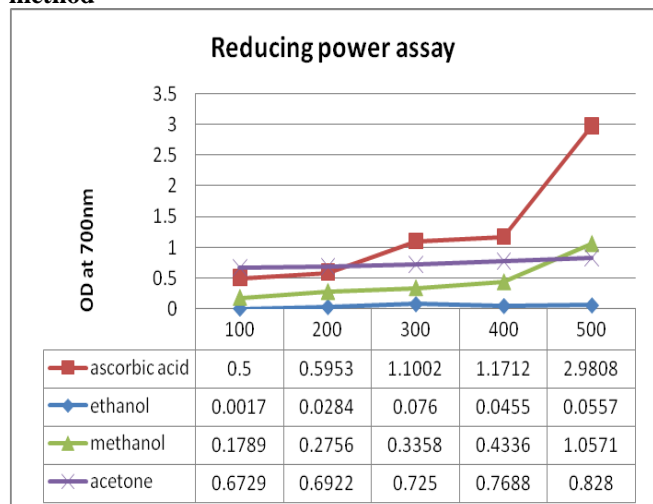


Fig 2. Different fractions tested by phospho-molybdenum assay

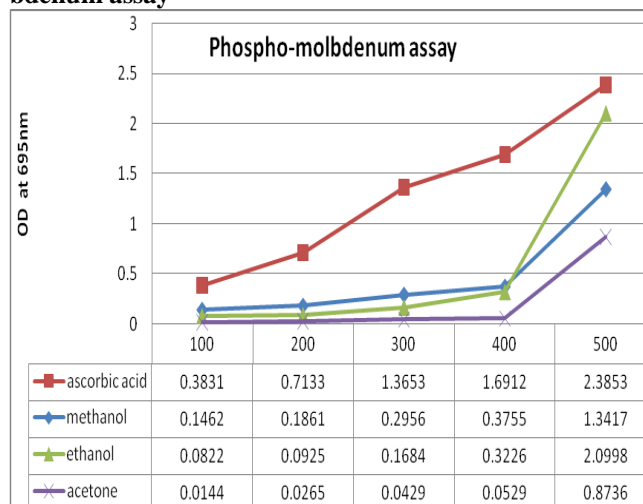
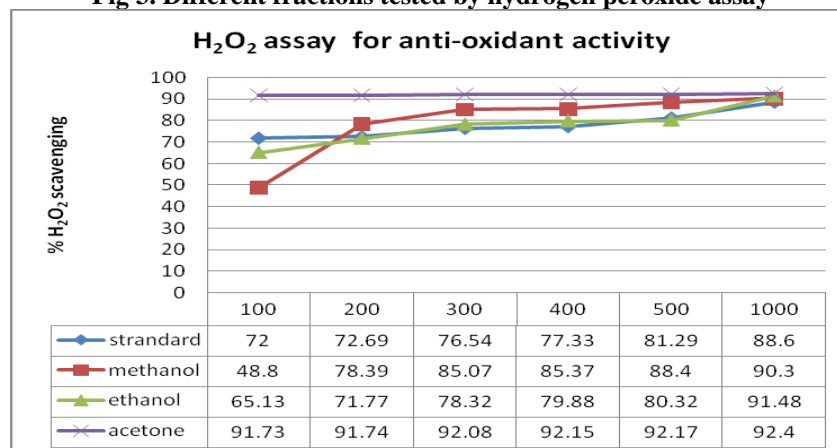


Fig 3. Different fractions tested by hydrogen peroxide assay**CONCLUSION**

Herbal drugs are being looked as very importance source for discovery of new agents for treating various ailments. *Lepidium sativum linn* showed good activity in scavenging the free radicals. Further pharmacological and clinical studies are required to understand the actual

efficacy of these herbal extracts

ACKNOWLEDGEMENT

Authors are thankful to the Oxford College of Science for providing laboratory facilities to carry out this research work

REFERENCES

- Divanji M, Vishwanath GL, et al. Ethnopharmacology of *Lepidium Sativum* Linn (Brassicaceae): A Review. *International Journal of Phytotherapy Research*, 2(1), 2012, 1-7.
- Nita D, Raval and Pandya TN *et al.*, Pharmacognostic study of *Lepidium sativum* Linn (Chandrashura). *Ayu*, 32(1), 2011, 116–119.
- Chanda S and Dave R. *In vitro* models for antioxidant activity evaluation and some medicinal plants possessing antioxidant properties: An overview. *African Journal of Microbiology Research*, 3(13), 2009, 981-996.
- Suvarchala Reddy NVL, Sneha JA and Raghavendra NM. *In Vitro* Antioxidant and Antidiabetic activity of *Asystasiagangetica* (Chinese Violet) Linn. (Acanthaceae). *International Journal of Research in Pharmaceutical and Biomedical Sciences*. 1(2), 2010, 72-75.
- Ashwini M, Nisha L, Shivaji B *et al.*, *In-vitro* antioxidant and anti-inflammatory activation activity of *Cocciniagrandsis*. *Int Jl. Pham. Pharm Sci*, 14 (3), 2012, 239-242.
- Roopashree TS, Raman D *et al.*, Antibacterial Activity Of Antipsoriatic Herbs: *Cassia Tora*, *Momordica Charantia* And *Calendula Officinali*. *International Journal Of Applied Research In Natural Products*, 1(3), 2008, 20-28.
- Pragya B, Dinesh B *et al.*, *In Vitro* Antioxidant Activity and Phytochemical Analysis Of Seed Extracts Of *Lepidium Sativum*. A Medicinal Herb. *J Biosci Tech*, 2(6), 2011, 410- 415.
- Hertog MGL, Feskens EJM *et al.*, Dietary antioxidant flavonoids and risk of coronary heart disease. The Zutphen elderly study. *The Lancet*, 342,(8878), 1993, 1007- 1014.
- Prasad AK, Kumar V, Arya P, Kumar S, Dabur R, Singh N, Chhillar AK, Sharma GL, Ghosh B, Wengel J, Olsen CE & Parmar VS. Investigations toward new lead compounds from medicinally important plants: *Pure Applied Chemistry*, 77(1), 2005, 25-40.
- Prashant T, Bimlesh K, Mandeep K, Gurpreet K, Harleen K. Phytochemical screening and Extraction: A Review. *International pharmaceutica scientia*, 1(1), 2011, 98-106.
- Kinnula VL, Crapo JD. Superoxide dismutases in malignant cells and human tumors. *Free Rad. Biol. Med*, 36, 2004, 718-744.
- Bergendi L, Benes L, Durackova Z, Ferencik M. Chemistry, physiology and pathology of free radicals. *Life Sci*, 65, 1999, 1865–1874.
- Prieto P, Pineda M and Aguilar M. Spectrophotometric quantitation of antioxidant capacity through the formation of phosphomolybdenum complex: Specific application to the determination of vitamin E. *Anal. Biochem*, 269, 1999, 337-341.
- Halliwell B. How to characterize an antioxidant: an update. *Biochem. Soc. Symp*, 61, 1995, 73-101.
- Aruoma OI. Free radicals, oxidative stress and antioxidants in human health and disease. *Jl. Am. Oil. Che. Soc.*, 75(2), 1998, 199-212.