



THE CONSTITUENTS AND BIOLOGICAL EFFECTS OF *ARUNDO DONAX* - A REVIEW

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ABSTRACT

Arundo donax is considered as one of the medicinal plants. Traditionally, the rhizome or rootstock was used for dropsy. Root or rhizome boiled in wine with honey was used for cancer. *Arundo donax* was also used for condylomata and indurations of the breast. The root infusion was considered as antilactagogue, depurative, diaphoretic, diuretic, emollient, hypertensive, hypotensive, and sudorific. *Arundo donax* is also used as haemostatic, in toothache, and for the treatment of pertussis and cystitis. It contained proteins, carbohydrates, lignin, cellulose, hemicelluloses, alkaloids, indoles, fatty acids, trace elements and many other bioactive metabolites. The previous studies showed that the plant exerted antibacterial, antifungal, anthelmintic, anticancer and other pharmacological effects. It also enhanced milk production and fattening performance. This review will highlight the constituents and pharmacological effects of *Arundo donax*.

Keywords: *Arundo donax*, traditional uses, chemical constituents, pharmacology.

INTRODUCTION

Plant derivatives had been employed by populations to prevent different kind of diseases for centuries. The knowledge of plant properties was acquired by ancient civilization that passed down from generation to generation until today. *Arundo donax* contained proteins, carbohydrates, lignin, cellulose, hemicelluloses, alkaloids, indoles, fatty acids, trace elements and many other bioactive metabolites. The previous studies showed that the plant exerted antibacterial, antifungal, anthelmintic, anticancer and other pharmacological effects. It also enhanced milk production and fattening performance.

Synonyms

Although there are many scientific names, but the currently accepted scientific name of giant reed is *Arundo donax* L. There are several recognized varieties, including *Arundo donax* var. *lanceolata* Döll; *Arundo donax* var. *variegata* E.Vilm.; *Arundo donax* f. *versicolor* (Mill.) Beetle; *Arundo donax* var. *angustifolia* Döll; *Arundo donax* var. *barbigera* (Honda) Ohwi; *Arundo donax* var. *coleotricha* Hack.; *Arundo donax* var. *versicolor* (Mill.) Kunth; and *Arundo donax* var. *versicolor* (Mill.) Stokes [1-3].

Taxonomic classification

Kingdom: Plantae

Phylum: Spermatophyta

Subphylum: Angiospermae

Class: Monocotyledonae

Order: Cyperales

Family: Poaceae

Genus: *Arundo*

Species: *Arundo donax* [2].

Common names

Arabic: kasab, zal, pos, ghab farsi, ghab roomi, kasab al-reeh, qish; Australia: bamboo, Danubian reed, e-grass, elephant grass, giant Danube grass, oboe reed; Brazil: cana do brejo, cana do reino, cana-brava, canno do reino, capim plumoso, taquara-do-reino; Chile: cañamo; Colombia: caña brava; Costa Rica: caña hueca; Cuba: caña de Castilla, caña de río, caña hueca, cañita de la india; Dominican Republic: cañita; English: Giant reed, Spanish reed, giant cane and giant feather-reed grass; Fiji: ngasau ni valangi; French: canne de Provence, grand roseau; Germany: Pfahlrohr; Haiti: herbe roseau, Roseau; Hindi: baranal, baru, doka, nal, naldura, nar, nara, narasal, narhal, narr, rajal, tinta; Italy: canna commune, canna comune, canna di Provenza; Kannada: baalada kaddi, bilee laalada kaddi, hulagilu hullu, hulugalagu, hulugila-hullu, hulugilu, hulugilu hullu, laalada kaddi; Malayalam: ama; Marathi: nal; Netherlands: Pijlriet; Puerto Rico: caña gigante, guajana; Sanskrit: dhamana, nala, potagala, sarah, sunyamadhya; South Africa: Spaanse-riet;

Spain: falso bamboo, gallipato alcublano, junco gigante; Tamil: caravanam, koraikkuccu, korukachi, korukkai, kuruvi-p-pul, velam; Telugu: adavikikasa-gaddi, adavikikasagadi, kaki veduru, paataveduru, patu veduru, peepalu; Tonga: kaho, kaho folalahi; Uruguay: caña musical [2-5].

Description

Arundo donax is a tall, erect, perennial cane- or reed-like grass. It was one of the largest herbaceous grasses, it can grow to 2-10 m tall. Its root structure is very strong, with the fleshy, almost bulbous, creeping rootstocks (rhizomes) forming compact bundles from which grow the fibrous roots, penetrating deep into the soil. The horizontal rhizomes give rise to many-stemmed, hollow, cane-like clumps allowing it to form large colonies many meters across. These tough, individual stems or culms are divided by partitions at the nodes like in bamboo, each node 12-30 cm in length and can reach diameters of 1-4 cm with walls 2-7 mm thick. They commonly branch during the second year of growth, rarely multiple, just single lateral branches from nodes. The outer tissue of the stem is of a siliceous nature, hard and brittle with a smooth glossy surface that turns pale yellow when the culm is fully mature. The pale, blue-green leaves clasp the stem broadly with a heart-shaped, hairy-tufted base, 2-6 cm wide at the base and tapering to a fine tip, up to 70 cm or more in length. The leaves are arranged alternately throughout the culm and very distinctly two-ranked, in a single plane. The culms can remain green throughout the year but often fade with semi-dormancy during the winter or in droughts. The flowers are borne in large plume-like panicles, 30-65 cm, at the upper tips of stems between March and September and are closely packed in a cream to brown-coloured cluster. The spikelets, flowering units comprised of one or more florets enclosed by two bracts or glumes, are several flowered, approximately 12 mm long with florets becoming successively smaller. The segmented central axis of the spikelet, the rachilla, is glabrous and dis-articulates above the glumes and between the florets. The more or less unequal glumes are 3-nerved membranous, narrow, slender, pointed and as long as the spikelets. Lemmas, the larger, outer, bract which, along with the palea, serves to contain the florets held within, are thin, 3-nerved and covered with fine, soft hair. They are narrowed upwards with the nerves ending in slender teeth [3,6-9].

Distribution

It was native to the Mediterranean area, introduced to many subtropical and warm temperate regions, where it was grown as an ornamental and was often found as a stray from cultivation [9-11]. It has been cultivated throughout Asia, southern Europe, northern Africa and the Middle East for thousands of years. In North America, it was intentionally introduced from the Mediterranean to the Los Angeles area in the early 1800s [12-14].

Traditional uses

The uses of *Arundo donax* have been dated back

to 5,000 B.C where the Egyptians used *Arundo donax* leaves as lining for underground grain storage. It has been stated that mummies were wrapped with *Arundo donax* leaves. In the 4th Century A.D. However, in more recent times *Arundo donax* has been commercially cultivated for the production of reeds for musical instruments. This type of commercial cultivation has been traced back for nearly 5,000 years. Not only has *Arundo donax* been used for musical instruments it has been cultivated for horticultural purposes, such as garden fences and trellises [15]. Medicinally, the rhizome or rootstock was used for dropsy. Root or rhizome boiled in wine with honey and used for cancer. *Arundo donax* was also used for condylomata and indurations of the breast. The root infusion was used as antigalactagogue, depurative, diaphoretic, diuretic, emollient, hypertensive and sudorific [16-20]. *Arundo donax* is also used as haemostatic, in toothache [19] and for the treatment of pertussis and cystitis [20].

Part used: rhizome, root leaves and stem were used medicinally

Physicochemical properties

Ash: 2.39%, extractives, cold water: 6.15%, hot water: 8.01%, sodium hydroxide 1%, alcohol-benzene: 1.29%, dichloromethane: nodes 13.04% and internodes 11.16%, ethanol: nodes 5.88% and internodes 4.18% [21-22].

Chemical constituents

Per 100 g, the green roughage is reported to contain on a zero moisture basis, 6.9 g protein, 1.3 g fat, 82.6 g total carbohydrate and 37.5 g fiber. The carbohydrates of *Arundo donax* consisted of: rhamnose 0.15 %, mannose 0.35 %, arabinose 2.00 %, galactose 0.84 %, xylose 33.88 %, glucose 62.13 %, 4-O-methyl glucuronic acid 0.65% [23]. *Arundo donax* showed an average of 22% lignin [21], 42% α -cellulose and 27% hemicelluloses. However, Shatalov *et al.*, observed that the levels of monosaccharides in the nodes and internodes (% respectively) were: arabinose 3.22 and 35.91, xylose 0.16 and 1.17, mannose 59.54 and 2.47, galactose 30.92 and 0.88 and glucose 0.83 and 64.91 [22]. It was also contained calcium 0.30–0.67%, P 0.08–0.15g, K 2.04–3.19%, and Mg 0.20–0.30%. The alkaloids especially gramine were isolated from the plant. Reed of Chinese origin contains 50.3% cellulose and 15.7% lignin [24-25].

The plant contained alkaloids including tryptamine, bufotenidine, gramine and arundamine [22,26]. Some studies have found that this plant was rich in active tryptamine compounds, Dutta and Ghosal were recovered 20 mg of DMT from 200 grams of the dry whole plant. It was accompanied by 520 mg of gramine, 128 mg of bufotenine and traces of 5-MeO-DMT, as well as unidentified indoles. Five indole-3-alkylamine bases, N,N-dimethyl-tryptamine, 5-methoxy-N-methyltryptamine, bufotenine, dehydro-bufotenine, and bufotenidine were isolated from the rhizomes of *Arundo donax* L [27-31].

The results of general chemical analysis evidenced the different relative abundance of holocellulose, lignin, proteins, extractives and ashes in internodes, nodes and foliage, at different stages of maturity. In internodes, nodes and foliage about 70–80% of hemicelluloses were easily extracted with 5% KOH aqueous solutions. The analysis of hemicelluloses indicated that they were consisted mainly from arabinoglucuronoxylans with a xylose, (arabinose: uronic acid ratio of 91–93:5–7:2 for internodes). The high content of xylose was found in *Arundo donax* stem. The easy extraction of hemicelluloses opens new perspectives for the use of the reed as a source of pentosans. The results obtained by the permanganate oxidation method indicated that *Arundo donax* lignins were essentially H-G-type with approximate H:G:S proportions of (32–36):(59–61):(5–8) in internodes. The H units are constituted mainly by esterified *p*-coumaric acid [32–34].

Milled wood lignin and dioxane lignin from different morphological regions (nodes and internodes) of *Arundo donax* reed were subjected to a comprehensive structural characterization. Both node and internode lignins are HGS-type lignins, with a significant amount of H units (including *p*-coumaric acid type structures). The S/G ratio was (1.13–1.32), the weight-average molecular weight was (20400–24500), the methoxyl group content was (0.90–0.98), the phenolic hydroxyl group content was (0.23–0.27), and the aliphatic hydroxyl group content was (1.00–1.09). However, some structural differences between node and internode lignins were observed. The former has much more phenolic acids (*p*-coumaric and ferulic), 8.8% in node versus 1.2% in internode and less β -O-4 (0.32 and 0.49 per aromatic unit) in node and internode, respectively [35–36].

Giant reed was low in protein but has a comparatively high concentration of phosphorus in the upper portions even when grown on soils with an extremely low concentration of this mineral. Mineral contents (percentages of oven-dry weight) included: calcium in the old plant, in the upper half 0.52 and in the lower half 0.67, in young plant in the upper half 0.30 and in the lower half 0.43; Magnesium in the old plant, in the upper half 0.25 and in the lower half 0.32, in young plant in the upper half 0.12 and in the lower half 0.19; Potassium in the old plant, in the upper half 2.04 and in the lower half 2.42, in young plant in the upper half 3.09 and in the lower half 3.19. Carbohydrates in the old plant, in the upper half 23.2 and in the lower half 21.7, in young plant in the lower half 20.0 and in the lower half 20.7 [37–38].

Chemical composition of lipophilic extractives in *Arundo donax* reed (mg/kg) included: *n*-Alkanes 77.9: *n*-Docosane 0.5, *n*-Tricosane 0.2, *n*-Tetracosane 0.6, *n*-Pentacosane 6.3, *n*-Hexacosane 3.9, *n*-Heptacosane 15.8, *n*-Octacosane 6.7, *n*-Nonacosane 37.0, *n*-Triacotane 0.8, *n*-Hentriacontane 5.4, *n*-Dotriacontane 0.3, *n*-Tritriacontane 0.4. Steroid hydrocarbons 127.4: Ergostatriene 14.5, Ergostadiene 9.3, Estigmastadiene 8.4, Estigmasta-3,5,22-triene 49.2, Estigmasta-3,5-diene 46.0. Fatty acids 1137.7: *n*-Tetradecanoic acid 3.5, *n*-

Pentadecanoic acid 1.8, *n*-Hexadecanoic acid 276.3, *n*-Heptadecanoic acid 10.0, 9,12-Octadecadienoic acid 30.0, 9-Octadecanoic acid 55.7, *n*-Octadecanoic acid 73.6, *n*-Nonadecanoic acid 3.1, *n*-Eicosanoic acid 50.0, *n*-Heneicosanoic acid 3.3, *n*-Docosanoic acid 35.7, *n*-Tricosanoic acid 25.3, *n*-Tetracosanoic acid 55.7, *n*-Pentacosanoic acid 33.5, *n*-Hexacosanoic acid 144.1, *n*-Heptacosanoic acid 14.3, *n*-Octacosanoic acid 134.9, *n*-Nonacosanoic acid 53.9, *n*-Triacotanoic acid 7 109.9, *n*-Hentriacontanoic acid 6.2, *n*-Dotriacontanoic acid 16.9. Fatty alcohols 194.3: *n*-Hexacosanol 33.4, *n*-Octacosanol 54.9, *n*-Triacotanol 57.7, *n*-Dotriacontano 48.3. Aldehydes 81.6: *n*-Hexacosanal 10.4, *n*-Octacosanal 22.9, *n*-Triacotanal 48.3. Sterols/triterpenols 528.1, Campesterol 90.6, Stigmasterol 46.4, B-Sitosterol 281.0, Stigmastanol 71.9, 7-oxo-Sitosterol 428 6.5, β -Amyrin 426 8.2, α -Amyrin 23.5. Tocopherol 17.7: β -Tocopherol 6.8, α -Tocopherol 10.9. Triterpenoid and steroid ketones 43.9: B-Amyrenone 10.2, α -Amyrenone 5.9, Cycloartenone 14.2, Stigmasta-3,5-dien-7-one 3.2, Stigmast-4-en-3-one 4.6, Stigmast-4-en-3,6-dione 3.6, Stigmastane-3,6-dione 2.5. Steryl/triterpenyl esters 68.1: Sitosteryl ester 16.1, B-Amyrinyl ester 14.0, α -Amyrinyl ester 38.0. Steryl glucosides 151.6: Campesteryl 3- β -d-glucopyranoside 30.6, Stigmasteryl 3- β -d-glucopyranoside 8.0, Sitosteryl 3-B-d-glucopyranoside 113.0. Monoglyceride 367.5: 2,3-Dihydroxypropyl tetradecanoate 5.5, 2,3-Dihydroxypropyl hexadecanoate 94.2, 2,3-Dihydroxypropyl octadecanoate 86.6, 2,3-Dihydroxypropyl eicosanoate 35.1, 2,3-Dihydroxypropyl docosanoate 43.0, 2,3-Dihydroxypropyl tetracosanoate 46.9, 2,3-Dihydroxypropyl hexacosanoate 56.2. Diglycerides 47.6: Dipalmitin, 1,2-(P2) 7.8, Dipalmitin, 1,3-(P2) 12.1, Palmitoylstearin (PS) 16.8, Distearin, 1,2- and 1,3-(S2) 10.9 [39].

PHARMACOLOGICAL EFFECTS

Antibacterial and antifungal effects

Aqueous extract of the stem nodes of *Arundo donax* exerted antibacterial activity against methicillin resistant *Staphylococcus aureus* (MRSA) in a concentration of 128 μ g/ml. The aqueous extracts of the reed nodes (which contain the white hemicellulose membrane) demonstrated a marked dose-dependent anti-biofilm activity, in preventing MRSA biofilm formation and disrupting established biofilms. These results may suggest that the traditional application of the reed membrane to fresh lacerations may be useful as a prophylactic for biofilm-related infection [40].

The antimicrobial effects of methanolic extracts of 14 medicinal plants species were examined comparing to conventional therapeutic antibiotics against standard bacterial strains (*Staphylococcus aureus*, *Micrococcus luteus*, *Klebsiella pneumonia*, *Escherichia coli* and *Pseudomonas aeruginosa*). *Arundo donax* extract showed the maximum effect against *Escherichia coli* and *Pseudomonas aeruginosa* among the examined fourteen medicinal plants species. The antimicrobial effects of 4% methanolic extracts of *Arundo donax* were comparable to Cephalotin (30mcg), Piperacilin (30mcg) and Amikacin

(30mcg) against *Escherichia coli* and Piperacilin (30mcg) and Amikacin (30mcg) against *Pseudomonas aeruginosa* [41].

Arundo donax also exerted antifungal activity against four Basidiomycetes (*Trametes versicolor* CTB 863A, *Coniophora puteana* BAM Ebw.15, *Gloeophyllum trabeum* BAM Ebw. 109, and *Postia placenta* FPRL 280) [21].

Anthelmintic effects

Crude aqueous-methanol extracts of the leaves of *Arundo donax* and its chloroform, petroleum spirit and ethyle acetate fraction were tested against *H. contortus*. *Arundo donax* (25-50 mg/ml) exerted anthelmintic effects ($P \geq 0.05$). 56.7% mortality of *H. contortus* was recorded by 10 hours post-exposure with crude aqueous methanol extracts of *Arundo donax* 50 mg/ml. There was 100% mortality of worms in Levamisole treatment (used as a reference drug) within 2 hours post-exposure. However, the anthelmintic effects of the plant was dose and time dependent. The ranking of efficacy of the *Arundo donax* fractions against *H. contortus* were ethyl acetate, chloroform aqueous followed by petroleum spirit fraction. Dose and time dependent ovicidal effects were recorded for these plant extracts. In egg hatch test, *Arundo donax* exhibited ovicidal activity with LC_{50} of 200.1 $\mu\text{g/ml}$; whereas, crude powder of *Arundo donax* resulted in 50.5% reduction in fecal egg count in sheep naturally infected with gastrointestinal nematodes [42].

Arundo donax extracts had anthelmintic properties (around 55% efficacy) against gastrointestinal parasites (*Ascaris* sp., *Oesophagostomum* sp. and *Paramphistomum* sp.) of cattle [43].

Antifeedant activity

Arundo donax revealed significant antifeedant activity against the boll weevil, *Anthonomus grandis* [31]. Bioassays indicated that while sterols showed limited antifeedant activity, significant activity was shown by the isolates tricontanol, tricin, and tetramethyl-N, N-bis-2,6 dimethylphenyl cyclobutane-1, 3-diimine [16,44].

Antiproliferative effect

Arundo donax was used in combination with *Spartium junceum* L. and *Cynodon dactylon* L. for the treatment of tumors (without specifying which kind of tumour) [45].

A lectin with antiproliferative activity towards human cancer cell lines and mitogenic towards human peripheral blood mononuclear cells was purified from the rhizomes of *Arundo donax* (Linn.). The molecular mass of native lectin was 32 kDa as determined by gel filtration chromatography. The *Arundo donax* lectin (ADL) was thermostable upto 55 °C and showed optimum activity in the range of pH 7.0–9.0 and comprised of 2.1% carbohydrate content [46].

A lectin (ADL) isolated and purified from *Arundo donax* rhizomes agglutinated native rabbit, pig erythrocytes and with lower intensity rat and human A, B and AB erythrocytes, and its hemagglutinating activity is

independent of divalent cations, but it was decreased by denaturing and reducing agents. *Arundo donax* lectin displayed cytotoxic effect on *Dysdercus peruvianus* and nematocidal activity against *Meloidogyne incognita*. ADL decreases the germinability and delayed the mean time for germinability of *Lactuca sativa* L. diaspores, and also showed significant mitogenic and chemotactic effect. The lectin induced toxicity in mice when administered intraperitoneally with dose of 300 mg/kg and 800 mg/kg, it caused 100 % death of the animals 30 h after its administration. Seven isoforms of ADL were separated. ADL-III is rich in Glu/Gln, Gly and Asp/Asn and Cys residues, and its N-terminal a and b chains contain tryptophan residues. ADL-III showed significant mitogenic activity. ADL was able to bind to transformed cells from T-47D, HT-29 and T-24 lines *in vitro*. Immunohistochemical techniques showed that ADL was localized in the fiber cell walls and in some few cortical parenchyma cells of the rhizome [47].

Effect on milk production and fattening performance

Due to its high fiber and low protein content, *Arundo donax* foliage is poorly digested. The reported digestibility values were about 47 to 52% for DM digestibility and about 54-56% for OM digestibility [48-51]. A commercial bolus made of a mixture of several powdered plants including *Arundo donax* L improved milk yield in dairy cows, which was attributed to the presence of components reported to be galactogogues [48-49]. However a higher DM digestibility of 69% was reported in an early study [52]. In lambs, fresh giant reed forage gave better fattening performance than giant reed hay [53]. Fresh giant reed forage and giant reed silage (ensiled 3% molasses on fresh basis) were better digested by Rahmani sheep than giant reed hay or berseem hay [51].

Central nervous effect

The rhizome of *Arundo donax* contained at least five tryptamines, including N,N-DMT, 5-MeO-DMT and bufotenine. It was reported that they produced no psychoactive effects [54], 50 mg of rhizome extract did not produce psychedelic effects [55], but instead, it caused mild but long lasting allergic reactions, which include blurred vision, watery and swollen eyes, conjunctivitis, and hives [54].

Effects on smooth muscles

A defatted ethanolic extract of the rhizomes produced hypotensive and antispasmodic effects against histamine-, serotonin- and acetylcholine- induced spasms. Bufotenidine showed three main pharmacological actions, antiacetylcholine effect, histamine release and uterine stimulant effects [56]. The alkaloid gramine was said to be a vasopressor, raising the blood pressure in dogs after small doses and causing a fall in larger doses [24,31,37,57-58].

Other pharmacological effects

Bufotenidine isolated from *Arundo donax* showed neuromuscular blocking activity [59]. *Arundo donax* L.,

was used as part of herbal preparation as herbal expectorant.

Dosage

Root 50-100 ml decoction [57].

CONCLUSION

Arundo donax exerted many pharmacological effects. There is a great promise for development of novel drugs from *Arundo donax* to treat human diseases.

REFERENCES

1. Lansdown RV. *Arundo donax*. The IUCN Red List of Threatened Species. *Arundo donax*, Version 2014.2. <http://www.iucnredlist.org/details/164340/0> [31 August 2014].
2. Invasive Species Compendium, *Arundo donax*, <http://www.cabi.org/isc/datasheet/1940> [6 August 2014].
3. Heuzé V, Tran G, Giger-Reverdin S and Lebas F. Giant reed (*Arundo donax*). Feedipedia.org. A programme by INRA, CIRAD, AFZ and FAO. <http://www.feedipedia.org/node/502> [15 August 2014, 18:37]
4. Shepherd RCH, Richardson RG and Richardson FJ. Plants of importance to Australia-a checklist, RG and FJ Richardson, Meredith, Victoria, 2001.
5. Douthit S. *Arundo donax* in the Santa Ana River Basin. In: Jackson, N. *Arundo donax* Workshop, 1994, 7-10.
6. Bell GP. Ecology and management of *Arundo donax*, and approaches to riparian habitat restoration in Southern California. In Wade JH, Pysek P, and Green D. (eds.), Plant Invasions: Studies from North America and Europe. Blackhuys Publishers, Leiden, The Netherlands, 1997.
7. Hitchcock AS and Chase A. Manual of the grasses of the United States. Misc Publ. 200. US Dept. of Agriculture, Washington DC, 1950, 184-185.
8. Duke JA. *Arundo donax* L. In: Handbook of Energy Crops, 1983.
9. Neal MC. In Gardens of Hawai'i. Bernice P. Bishop Museum Special Publication 40, Bishop Museum Press, Honolulu, HI, 1965.
10. Wagner WL, Herbst DR and Sohmer SH. Manual of the Flowering Plants of Hawai'i. 2 vols. Bishop Museum Special Publication 83, University of Hawai'i and Bishop Museum Press, Honolulu, Hawaii, 1999.
11. Hoshovsky M. Element stewardship abstract: *Arundo donax* - giant reed, [Online]. In: Invasives on the web: The Nature Conservancy wildland invasive species program. Davis, CA: The Nature Conservancy, 1989.
12. Dudley TL. *Arundo donax* L. In: Bossard, Carla C, Randall, John M, Hoshovsky, Marc C., eds. Invasive plants of California's wildlands. Berkeley, CA: University of California Press, 2000, 53-58.
13. Csurhes S. Weed Risk Assessment: Giant reed (*Arundo donax*). The State of Queensland, Department of Employment, Economic Development and Innovation, Australia, 2009.
14. Oakins AJ. An Assessment and management protocol for *Arundo donax* in the Salinas Valley Watershed. Bachelor thesis, California State University, 2001.
15. Miles DH, Tunsuwan K, Chittawong V, Hedin PA, Kokpol U, Ni CZ and Clardy J. Agrochemical activity and isolation of N-(bromophenyl)-2,2-diphenylacetanilide from the Thai plant *Arundo donax*. *J Nat Prod*, 56, 1993, 1590-1593.
16. Bandaranayake WM. Bioactivities, bioactive compounds and chemical constituents of mangrove plants. *Wetlands Ecology and Management*, 10, 2002, 421-452.
17. Duke JA and Wain KK. Medicinal plants of the world. Computer index with more than 85,000 entries. 3, 1981.
18. Passalacqua NG, Guarrera PM and De Fine G. Contribution to the knowledge of the folk plant medicine in Calabria region (Southern Italy). *Fitoterapia*, 78, 2007, 52-68.
19. Safa O, Soltanipoor MA, Rastegar S, Kazemi M, Dehkordi KN and Ghannadi A. An ethnobotanical survey on hormozgan province, Iran. *Avicenna Journal of Phytomedicine*, 3(1), 2003, 64-81.
20. Temiz A, Akbas S, Panov D, Terziev N, Hakk M, Parlak S and Kose G. Chemical composition and efficiency of bio-oil obtained from giant cane (*Arundo donax* L.) as a wood preservative. *Bio Resources*, 8(2), 2013, 2084-2098.
21. Shatalov AA, Quilhó T and Pereira H. *Arundo donax* L. reed: new perspectives for pulping and bleaching raw material characterisation. *Tappi Journal Review Edpaper*, 84 (1), 2001, 1-12.
22. Miller DF. Composition of cereal grains and forages. National Academy of Sciences, National Research Council, Washington, DC Publ, 1958, 585.
23. Orlandi M, Luca Z and Salanti A. Characterization of lignocellulosic materials during the biorefinery process of *Arundo donax* for fine chemicals production. University of Milano-Bicocca, Department of Earth and Environmental Sciences, 2013.
24. Gohl B. Tropical feeds. Feed information summaries and nutritive values. FAO Animal Production and Health Series 12. FAO, Rome, 1981.
25. Erowid *Arundo donax* Info Page 1 and 3, <https://www.erowid.org/search.php?exclude=&q=Arundo+donax&x=14&y=3> [Oct 2012].
26. Dutta SK and Ghosal S. Indole-3-alkylamines of *Arundo donax* L. *Chemistry and Industry*, 1967, 2046-2047.
27. Ghosal S. Occurrence of psychedelic substances in some Indian medicinal plants. *Planta Medica*, 21(2), 1972, 200-209.
28. Ghosal S, Chaudhuri RK and Dutta SK. Alkaloids of the flowers of *Arundo donax* L. *Phytochemistry*, 10, 1971, 2852-2853.

29. Ghosal S, Chaudhuri RK, Dutta SK and Bhattacharya SK. Occurrence of curaromimetic indoles in the flowers of *Arundo donax*. *Planta Med*, 1972, 21-22.
30. Ghosal S, Dutta SK, Sanyal AK and Bhattacharya SK. *Arundo donax*L. (Graminae). Phytochemical and pharmacological evaluation. *Journal of Medicinal Chemistry*, 12, 1969, 480-483.
31. Neto CP, Seca A, Nunes AM, Coimbra MA, Domingues F, Evtuguin D, Silvestre A and Cavaleiro JAS. Variations in chemical composition and structure of macromolecular components in different morphological regions and maturity stages of *Arundo donax*. *Industrial Crops and Products*, 6(1), 1997, 51-58.
32. Joseleau JP and Barnoud F. Hemicelluloses of young internodes of *Arundo donax*. *Phytochemistry*, 13, 1974, 1155-1158.
33. Joseleau JP and Barnoud F. Hemicelluloses of *Arundo donax* at different stages of maturity. *Phytochemistry*, 14, 1975, 71-75.
34. Seca AML, Cavaleiro JAS, Domingues FMG, Silvestre AGD, Evtuguin D and Neto CP. Structural characterization of the lignin from the nodes and internodes of *Arundo donax* reed. *J Agric Food Chem*, 48, 2000, 817-824.
35. Faix O, Meier D and Beinhoff O. Analysis of lignocelluloses and lignins from *Arundo donax* and *Miscanthus sinensis* Anderss and hydroliquefaction of *Miscanthus*. *Biomass*, 18, 1989, 109.
36. Perdue Jr, RE. *Arundo donax*-source of musical reeds and industrial cellulose. *Economic Botany*, 12, 1958, 368-404.
37. Wynd FL, Steinbauer George P and Diaz NR. *Arundo donax* as a forage grass in sandy soils. *Lloydia*, 11(3), 1948, 181-184.
38. Coelho D, Marques G, Gutierrez A, Silvestre AJD and delRio JC. Chemical characterization of the lipophilic fraction of giant reed (*Arundo donax*) fibres used for pulp and paper manufacturing. *Industrial Crops and Products*, 26, 2007, 229-236.
39. Quavea CL, Plano LRW, Pantuso T and Bennett BC. Effects of extracts from Italian medicinal plants on planktonic growth, biofilm formation and adherence of methicillin-resistant *Staphylococcus aureus*. *Journal of Ethnopharmacology*, 118, 2008, 418-428.
40. Shirvani A, Mozaffari M and Zarei M. Antimicrobial effects of 14 medicinal plant species of dashti in Bushehr province. *Iranian south Medical Journal (ISMJ)*, 17(1), 2014, 49-57.
41. Badar N. Documentation of indigenous antiparasitic practices and scientific evaluation of some ethnobotanicals for their anthelmintic activity. PhD thesis, Faculty of Veterinary Science, University of Agriculture Faisalabad, Pakistan, 2011
42. Sharatkumar S, Dhanachand C and Mohilal N. Study on the efficacy of certain medicinal plants on gastrointestinal helminths of cattle. *Indian Vet J*, 81(5), 2004, 497-498.
43. Miles DH, Tunswan K, Chittawong V, Kokpol U, Choudhary MI and Clardy J. Boll weevil antifeedants from *Arundo donax*. *Phytochemistry* 1993; 34(5): 1277-1279.
44. Leporatti ML and Impieri M. Ethnobotanical notes about some uses of medicinal plants in Alto Tirreno Cosentino area (Calabria, Southern Italy). *Journal of Ethnobiology and Ethnomedicine*, 3, 2007, 34-39.
45. Kaur A, Singh J, Kamboj SS, Sexana AK and Shamugavel M. Isolation of an N-acetyl-D-glucosamine specific lectin from the rhizomes of *Arundo donax* with antiproliferative activity. *Phytochemistry*, 66(16), 2005, 1933-1940.
46. Zanetti GD. (Lectina dos rizomas de *Arundo donax* L.: purificação, caracterização, propriedades, imuno-histoquímica e separação das isoformas) *Arundo donax* L. rhizomes lectin : purification, characterization, properties, immunohistochemistry and separations of isoforms. PhD thesis, Universidade Federal do Rio Grande do Sul. Instituto de Biociências. Programa de Pós-Graduação em Botânica, 2007.
47. Baig MI and Bhagwat VG. Study the efficacy of Galactin Vet bolus on milk yield in dairy cows. *Vet World*, 2(4), 2009, 140-142.
48. Behera PC, Tripathy DP and Parija SC. Shatavari: potentials for galactagogue in dairy cows. *Indian J Trad Knowledge*, 12(1), 2013, 9-17.
49. Tagel-Din AE. Evaluation of reeds in complete diets for ruminant animals. *Indian J Anim Sci*, 60(9), 1990, 1106-1109.
50. Ahmed ME, Abdel-Gawad AM, Shehata EI and Tawfik SA. Influence of using reed forage in different forms as fresh, silage and hay on blood profile and carcass quality of growing Rahmani sheep. *Egyptian J Sheep Goat Sci*, 6(2), 2011, 25-35.
51. Talapatra SK. The nutritive value of the indigenous grasses of Assam. III. The semi-aquatic grasses as cattle feeds. *Indian J Vet Sci*, 20, 1950, 229-240.
52. Ahmed M E, Shehata E I, Ammou FFA, Khalifa EI and El-Zolaky OA. Productive and reproductive performance of Rahmani sheep fed rations containing reed forage (*Arundo donax* L.) either fresh, hay or silage. *Egyptian J Sheep Goat Sci*, 4(1), 2009, 45-54.
53. DeKorne J. *Arundo donax*. *Entheogene*, 4, 1995, 27-28.
54. Christian R. The Encyclopedia of psychoactive plants: Ethnopharmacology and its applications. Rochester, Park Street Press, 1998.
55. Ana ML, Seca Jose AS, Cavaleiro FMJ, Domingues AJD, Silvestre DE and Carlos PN. Structural characterization of the lignin from the nodes and internodes of *Arundo donax*. *J Agric Food Chem*, 48, 2003, 817-824.
56. Khare CP. *Indian Medicinal Plants- An illustrated dictionary*. Springer Science and Business Media LLC, 2007, 66-67.
57. Chopra RN, Nayyar SL and Chopra IC. *Glossary of Indian medicinal plants*. Council of Scientific and Industrial Research, New Delhi, India 1956, 160.

58. Bhattacharya SK and Sanyal AK. Neuromuscular blocking activity of bufotenidine isolated from *Arundo donax* L. *Naturwissenschaften*, 59(12), 1972, 650-651.
59. Menale B, Amato G, Prisco D and Muoio R. Traditional uses of plants in North-Western Molise (Central Italy). *Delpinoa*, 48, 2006, 29-36.